

# ASN3<sub>50</sub> + B + N/2

(Modified from: Rippka R., 1988. Isolation and purification of cyanobacteria. *Method Enzymol.* 167: 3-27.)

Minerals	Stock Solutions (g/L)	Quantity (mL Stock/L Media)	Final Concentration (mM)
NaCl	250	100	428
MgCl <sub>2</sub> · 6H <sub>2</sub> O	200	10	10
KCl	50	10	6.5
MgSO <sub>4</sub> · 7H <sub>2</sub> O	350	10	14
CaCl <sub>2</sub> · 2H <sub>2</sub> O	50	10	3
Na <sub>3</sub> -citrate	0.6	5	0.012
Na <sub>2</sub> -EDTA · 2H <sub>2</sub> O	0.1	5	0.0013
Trace metal mix (A5 + Co)	See recipe below	1	-

Adjust to 900mL with mQ water and autoclave.

After cooling, add the following filter sterilized (0.2 µm) components to complete the medium:

Minerals	Stock Solutions (g/L)	Quantity (mL Stock/L Media)	Final Concentration (mM)
NaNO <sub>3</sub>	150	2.5	4.4
K <sub>2</sub> HPO <sub>4</sub> · 3H <sub>2</sub> O	4	5	0.088
NaHCO <sub>3</sub>	18	10	2
Fe-NH <sub>4</sub> -citrate	6	0.5	-
NH <sub>4</sub> Cl	53.5	0.5	0.5
Vitamin B12 (Cyanocobalamin)	0.02	1	-

For solid medium use 7g/L of agarose. Sterilize the agarose separately in 550 ml of milliQ water. In this case the mineral solution is filled up to 400 ml.

## Trace metal mix A5 + Co:

Trace metals	Quantity g/L	Concentration in the final media (mM)
H <sub>3</sub> BO <sub>3</sub>	2.86	0.047
MnCl <sub>2</sub> · 4H <sub>2</sub> O	1.81	0.009
ZnSO <sub>4</sub> · 7H <sub>2</sub> O	0.22	0.0007
Na <sub>2</sub> MoO <sub>4</sub> · 2H <sub>2</sub> O	0.39	0.0016
CuSO <sub>4</sub> · 5H <sub>2</sub> O	0.08	0.0003
Co(NO <sub>3</sub> ) <sub>2</sub> · 6H <sub>2</sub> O	0.05	0.0002